Optimizing silver nitrate concentration for enhanced Date Palm (*Phoenix dactylifera* L.) Tissue culture propagation and contamination management: A quantitative analysis

Yuvraj Sinh Vala¹, Hitesh Nakum², and Pradeep U Verma³

1&2 Research Scholar, Department of Botany, Shree Swaminarayan Science College, Faculty of Science, Swaminarayan University, Kalol, Gandhinagar, Gujarat.

³Dean and Professor, Department of Botany, Shree Swaminarayan Science College, Faculty of Science, Swaminarayan University, Kalol, Gandhinagar, Gujarat.

Abstract:

Date palm (*Phoenix dactylifera* **L.**) is a crucial crop in arid and semi-arid regions globally, prized for its fruit and resilience. However, in vitro propagation is often hindered by microbial contamination and suboptimal growth conditions. Silver nitrate (AgNO₃), renowned for its antimicrobial properties, has been employed as an effective agent in the tissue culture. This study investigates the optimal AgNO₃ concentration required for achieving a balance between contamination control and plant growth promotion in the date palm tissue culture. By analyzing a range of AgNO₃ concentration (0-625 μ g/L), we found that 250 μ g/L AgNO₃ yielded the highest performance reducing contamination to 13% promoting a shoot elongation to 5.6cm and achieving 85% root induction. Higher concentration (\geq 375 μ g/L) resulted in phytotoxicity and inhibited growth. The findings suggest that 250 μ g/L is the optimal concertation for maximizing tissue culture success and minimizing contamination in the date palm propagation.

Keywords: Silver Nitrate, Contamination Management Phytotoxicity, Date Palm Tissue Culture, *Phoenix Dactylifera*.

Introduction: Date palm hold the immense agricultural and economic significances, particularly in the arid regions. Its successful mass propagation relies on the in-vitro techniques, which are often compromised by the microbial contamination and variability in the growth responses.(1)(2)

Silver nitrate is known for its dual role in the mitigating microbial activity and modifying ethylene-mediated responses, making it a promising candidate for improving tissue culture outcomes.(3)(4)

This study aims to explore the effects of AgNO₃ in controlling contamination and promoting the developmental parameters in the date palm cultures. By systematically varying its concentration in the culture medium, we sought to establish its optimal dosage for maximal efficacy. From this it will establish the understanding of the concentration-dependent effects of AgNO₃ will help refine tissue culture protocols and promote more efficient and reproducible results.

Material and Methods

1. **Plant material**: Embryos and explants from off shoots of the date palm (Elite Kutch) cultivated in Bhuj, India. The samples underwent surface sterilization using a sequential treatment of 70% ethanol for 2 minutes, followed by 0.1% mercuric chloride for 10 minutes. After sterilization. Explants were thoroughly rinsed with sterile distilled water to remove any residual disinfectant. The explants were cultured on MS Medium supplemented with various concentration of AgNO₃ (0,125,250,375,625 μg/L). The medium was supplemented with 6-benzylaminopuine (BAP) as a growth regulator for shoot induction. Cultures were maintained in a growth chamber at 25°C ± 2°C under a 16-hour light/8-hour dark photoperiod.

2. Experimental design:

A *completely randomized design* (CRD) was adopted, with five treatments and three replications per treatment (n=3).(5). Each treatment contained 15 explants. The cultures were monitored for microbial contamination, shoot elongation, and root induction at 4,8, and 12 weeks. AgNO₃ was introduced into the culture medium at five concentrations: 0, 125, 250, 375, 500, and 625 μg/L. Two parameters were evaluated:

- 1. **Contamination-free cultures:** It was assessed by visual inspection of microbial growth, recorded as a percentage of contaminated cultures at 72 hours post culture initiation.
- 2. **Shoot elongation:** The length of the longest shoot was measured at 4 and 8 weeks to assess shoot growth.
- 3. **Root Induction:** The percentage of explants with developed roots was recorded at 8 and 12 weeks. The effectiveness index was calculated to determine the optimal AgNO₃ concentration using the formula. (6)

Effective index =
$$\frac{\text{Success rate} \times \text{Root induction rate}}{100 - \text{contamination rate}}$$

Statistical Analysis:

Data were analyzed using one-way ANOVA and t-test with the control group (0 ug/L) to identify significant differences between treatments. Post Hoc comparisons were conducted using the Tukey HSD test (p<0.05). All statistical analyses were performed using IBM SPSS 21.0 software

Results and Discussion

Microbial Contamination control: At 250μg/L AgNO₃ the concentration rate was reduced to 13%, which was significantly lower than the control group with 55% contamination. Higher AgNO₃ concentration (375μg/L and 625μg/L) resulted in increased contamination rates, likely due to phytotoxicity at these levels, which adversely affected explant health and compromised contamination control.(7)(8)

Shoot Elongation and root induction: Explants cultured with $250\mu g/L$ AgNO3 exhibited the highest shoot elongation (5.6cm) and 85% root induction. This concentration provided the most favorable conditions, promoting both shoot and root development without causing phytotoxicity. In contrast, higher AgNO3 concentration (375 $\mu g/L$ and above) resulted in significant growth inhibition, with reduced shoot elongation and root induction. The growth inhibitory effect of AgNO3 at these concentrations was evident by the stunted shoots and poor root development with root induction rates falling below 30% at 625 $\mu g/ml.(9)$

Effectiveness index: The effectiveness index was highest at $250\mu g/L$ AgNO₃ with a value of 4.87, indicating the optimal balance between contamination control and growth promotion. Concentrations above 375 $\mu g/L$ shown a reduction in the effectiveness index, highlighting the detrimental effects of higher AgNO₃ concentration on plant growth. (10)

Underlying Mechanism: The antimicrobial action of silver nitrate stems from its ability to disrupt the microbial cell ways and interfere with the DNA replication mechanism. Besides, its role in the modulating the ethylene responses may explain the observed enhancements in the plant morphogenesis, as the ethylene often inhibits the growth under in vitro conditions.(11)

This study highlights the concentration-dependent effects of silver nitrate (AgNO₃) on date palm tissue culture, demonstrating that 250µg/L AgNO₃ provides the optimal concentration for controlling the microbial contamination while simultaneously promoting growth. The antimicrobial properties of AgNO₃, are critical in maintaining sterile conditions necessary for successful tissue culture propagation.(12)

At concentration below $250\mu g/L$. AgNO₃ showed insufficient antimicrobial efficacy allowing contamination to persist. Conversely concentration above $375\mu g/L$ resulted in phytotoxicity, which manifested as stunted growth and inhibited root development consistent with the findings in other plant species (**Table 1**).

The mechanism of AgNO₃'s action appears to involve its ability to inhibit the microbial growth while maintain plant cell viability. This study also suggests that ethylene inhibition plays a significant role in promoting growth under AgNO₃ treatment. (13)

Ethylene, a plant hormone associated with the stress is known to inhibit the root and shoot development. By reducing ethylene action, AgNO₃ helps in optimal growth and regeneration.(14) (15) (Table 2)

Table 1

Effect of different Concentration of AgNO ₃ in the culture media					
	Contamination percentage				
AgNO ₃ concentration s (μg/L)	Establishment Stage	Callus stage	Multiplication & germination stage		
0	20.11	15.44	11.15		
125	15.44	11.15	10.31		
250	10.32	10.31	7.21		
375	7.21	7.21	4.32		
500	4.35	4.32	2.24		
625	2.26	2.24	0.00		

Callus induction $\% = \frac{number\ of\ explant}{total\ number\ of\ explants} \times 100$

Table 2

Effect of Silver Nitrate on callus stage					
AgNO ₃ CONCENTR ATIONS	CALLUS FORMATION %	CALLUS GROWTH	GLOBULAR EMBRYO FORMATION		
(µg/L)					
0	40.88	1.77	8.00		
125	45.55	2.66	17.00		
250	58.77	3.11	14.00		
375	62.66	3.44	12.00		
500	70.33	3.88	7.00		
625	74.44	4.22	8.00		

 $\textit{Direct embryo Percentage} = \frac{\text{number of explant forming direct embryo}}{\text{total number of embryo}} \times 100$

Table 3:-

Effect of Silver Nitrate on date palm during embryo stage					
AGNO ₃	Direct embryo	Embryo	Embryo		
CONCENTR	formation %	multiplication	germination		
ATIONS					
(μg/L)					
0	30.11	0.33	0.33		
125	35.22	0.48	0.50		
250	43.22	0.55	0.65		
375	48.22	0.68	0.72		
500	48.55	0.88	0.81		
625	68.11	1.07	0.95		

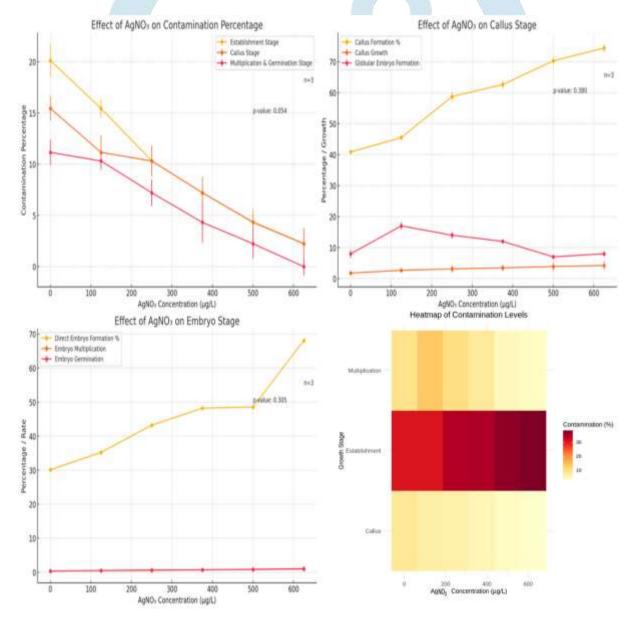


Fig.1: The contamination percentgaes at different AgNO₃ concentration (0-625ug/L) across the samples (n=3) tissue culture stages: establishment, callus, and mulitplication and germination. A significant reduction in the contamination was observed with the increasing concentrations, with the lowest contamination rates recorded at 625ug/l. Statitical analysis revealed that the differences between 250ug/l and higher concentration (375-625ug/L) was significant for contamination reduction, but higher concentration showed the potential phytotoxic effects as disccused in the text. **Fig.2:** Effect of AgNO₃ on the globular embryo formation in the date palm tissue culture-Illustrates the globular embryo formation percentages as influenced by AgNO₃ concentrations. The formation peaked at 125ug/l (17%) followed by a decline with higher concentrations. Statistical analysis (ANOVA, p<0.380) indicate that embryo formation at 125ug.L was significantly higher than the that at 500-625 ug/L, highlighting potential inhibitory effects of elevated AgNO₃ levels.

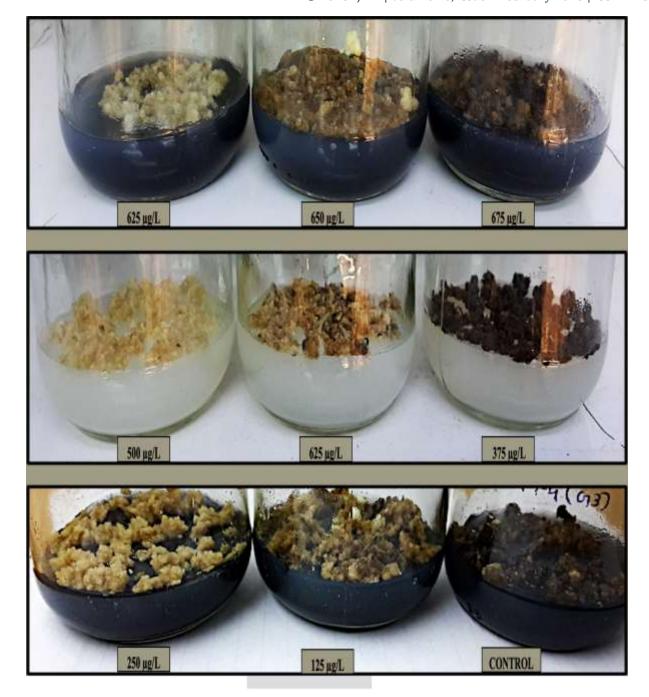


Fig. 3(A) Effect of AgNO3 on the induction ,growth and somatic embryogenesis germination of the Kutch elite date.

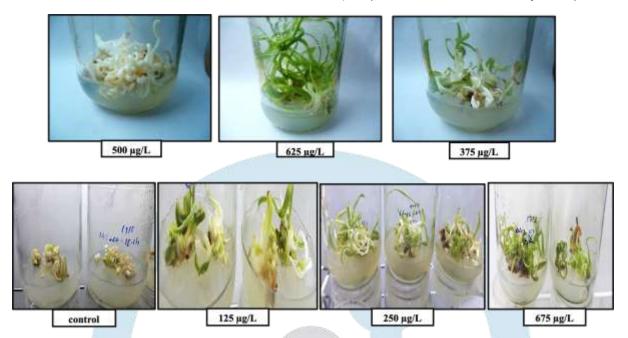


Fig .3 (B) Effect of AgNO3 on the embryo formation, mulitplication and somatic embryogenesis germination of the Kutch elite

Conclusion: The findings of this study indicates that 250µg/L AgNO₃ is the optimal concentration for the contamination control and growth promotion in date palm tissue culture. At this concentration microbial contamination was minimized, while shoot elongation and root induction were maximized. The results demonstrate that increasing the concentrations of AgNO₃ reduce the contamination rates and enhance the callus formation, growth and embryo development. The most significant improvements were observed at 625ug/L, with contamination percentage dropping to a significant lower stage. However, higher concentrations also led to a reduction in the globular embryo formation, indicating a potential inhibitory effect beyond a certain threshold

The results provide important insights for optimizing in vitro propagation protocols for the date palm ultimately improving the efficiency of propagation and enhancing the quality of plantlets produced.

Reference:

- 1. Jaradat, A.A., 2015. Biodiversity, genetic diversity, and genetic resources of date palm. *Date Palm Genetic Resources and Utilization: Volume 1: Africa and the Americas*, pp.19-71.
- 2. Jaradat, A.A., 2016. Genetic erosion of Phoenix dactylifera L.: perceptible, probable, or possible. *Genetic Diversity and Erosion in Plants: Case Histories*, pp.131-213.
- 3. Thao, N.P., Khan, M.I.R., Thu, N.B.A., Hoang, X.L.T., Asgher, M., Khan, N.A. and Tran, L.S.P., 2015. Role of ethylene and its cross talk with other signaling molecules in plant responses to heavy metal stress. *Plant Physiology*, *169*(1), pp.73-84.
- 4. Iqbal, N., Trivellini, A., Masood, A., Ferrante, A. and Khan, N.A., 2013. Current understanding on ethylene signaling in plants: the influence of nutrient availability. *Plant Physiology and Biochemistry*, 73, pp.128-138.
- Khafri, A.Z., Zarghami, R., Ma'mani, L. and Ahmadi, B., 2023. Enhanced efficiency of in vitro rootstock micro-propagation using silica-based nanoparticles and plant growth regulators in myrobalan 29C (Prunus cerasifera L.). *Journal of Plant Growth* Regulation, 42(3), pp.1457-1471.
- 6. He, F., Shi, C., Yuan, Q., Chen, C. and Zheng, K., 2012. AgNO3-based colorimetric methods for measurement of chloride penetration in concrete. *Construction and Building Materials*, 26(1), pp.1-8.
- 7. Prasad, A., Sidhic, J., Sarbadhikary, P., Narayanankutty, A., George, S., George, B.P. and Abrahamse, H., 2024. Role of metal nanoparticles in organogenesis, secondary metabolite production and genetic transformation of plants under in vitro condition: a comprehensive review. *Plant Cell, Tissue and Organ Culture (PCTOC)*, 158(2), p.33.
- 8. Zafar, H., Javed, R. and Zia, M., 2023. Nanotoxicity assessment in plants: an updated overview. *Environmental Science and Pollution Research*, 30(41), pp.93323-93344.
- 9. Saleeb, N., 2019. Interaction of Silver Nanoparticles and Silver Ions with Soil, Plant and Earthworm Aportectodea caliginosa: A dissertation submitted in partial fulfilment of the requirements for the Degree of Doctor of Philosophy at Lincoln University (Doctoral dissertation, Lincoln University).

- 10. Tripathi, A., Liu, S., Singh, P.K., Kumar, N., Pandey, A.C., Tripathi, D.K., Chauhan, D.K. and Sahi, S., 2017. Differential phytotoxic responses of silver nitrate (AgNO3) and silver nanoparticle (AgNps) in Cucumis sativus L. *Plant Gene*, 11, pp.255-264.
- 11. Pandian, S.R.K., Deepak, V., Kalishwaralal, K., Viswanathan, P. and Gurunathan, S., 2010. Mechanism of bactericidal activity of silver nitrate-a concentration dependent bi-functional molecule. *Brazilian Journal of Microbiology*, 41, pp.805-809.
- 12. Drisya Ravi, R.S., Siril, E.A. and Nair, B.R., 2019. The effect of silver nitrate on micropropagation of Moringa oleifera Lam. an important vegetable crop of tropics with substantial nutritional value. *Physiology and Molecular Biology of Plants*, *25*, pp.1311-1322.
- 13. Munkager, V., Vestergård, M., Priemé, A., Altenburger, A., de Visser, E., Johansen, J.L. and Ekelund, F., 2020. AgNO3 sterilizes grains of barley (Hordeum vulgare) without inhibiting germination—a necessary tool for plant–microbiome research. *Plants*, *9*(3), p.372.
- 14. Neves, M., Correia, S., Cavaleiro, C. and Canhoto, J., 2021. Modulation of organogenesis and somatic embryogenesis by ethylene: An overview. *Plants*, 10(6), p.1208.
- 15. Lee, J., Naing, A.H., Park, K.I. and Kim, C.K., 2023. Silver nitrate reduces hyperhydricity in shoots regenerated from the hypocotyl of snapdragon cv. Maryland Apple Blossom. *Scientia Horticulturae*, 308, p.111593.

